Emergence of *Campylobacter jejuni* ST-6964 in poultry and humans in New Zealand: a new twist in the campy story

French NP¹, Williamson DA^{2,3}

Biggs R⁴, Biggs PJ¹, Bloomfield S¹, Dyet K², Gilpin BJ², Heffernan H², Midwinter AC¹, Mulqueen K⁴, Rogers LE^{1,,} Wilkinson, D¹, Zhang, J¹

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One Health Aotearoa 2016, Wellington





Collaborating Centre



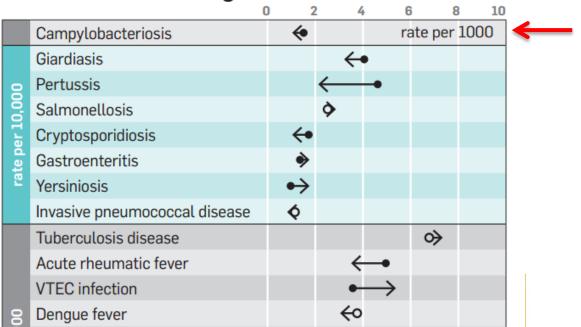


Campylobacteriosis still high on the list of notifiable diseases in New Zealand, but declining

New Zealand Public Health Surveillance Report September 2015: Covering April to June 2015

Source: ESR Ltd

National surveillance data 12-monthly notification rate changes¹

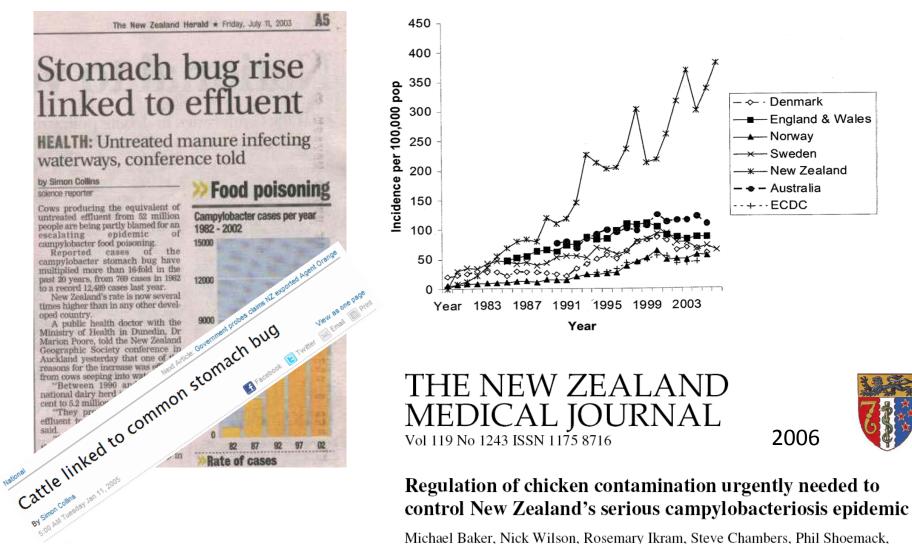




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Campylobacter in NZ: 1980-2006



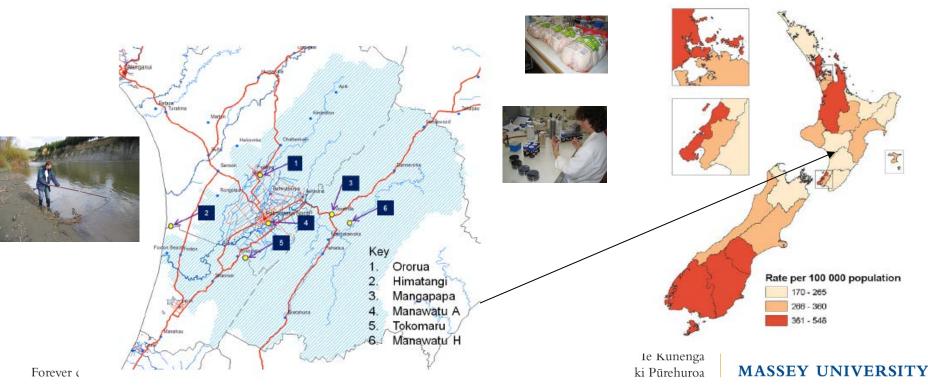
Gregory Cook

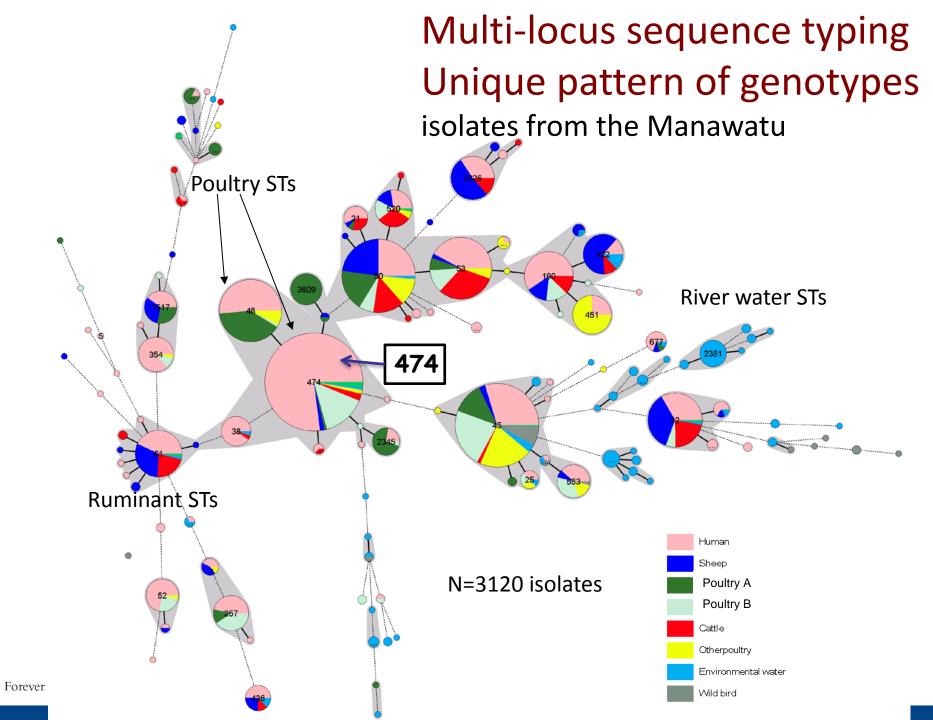
Manawatu sentinel site 2005-2015

- 'One health' approach
- >7000 samples
- Identify MLST types common to particular sources

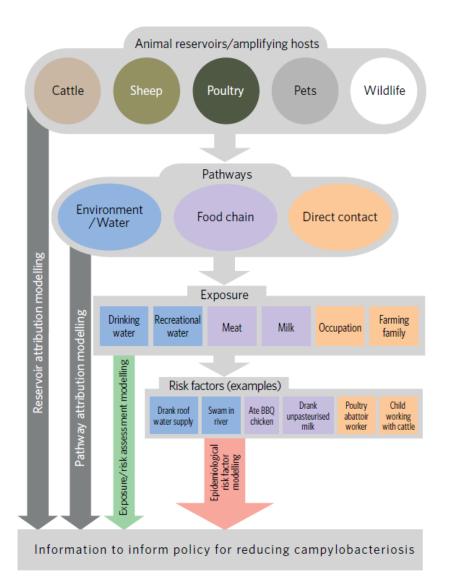
Figure 7. Campylobacteriosis notifications by DHB, 2004

• Modelling (reservoir attribution)

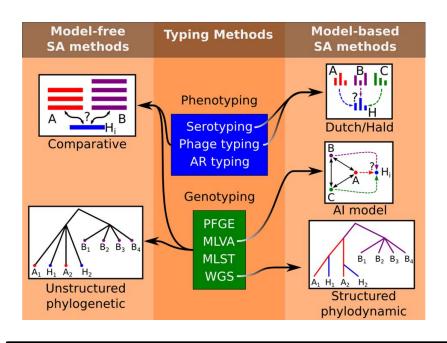




Source attribution and transmission frameworks



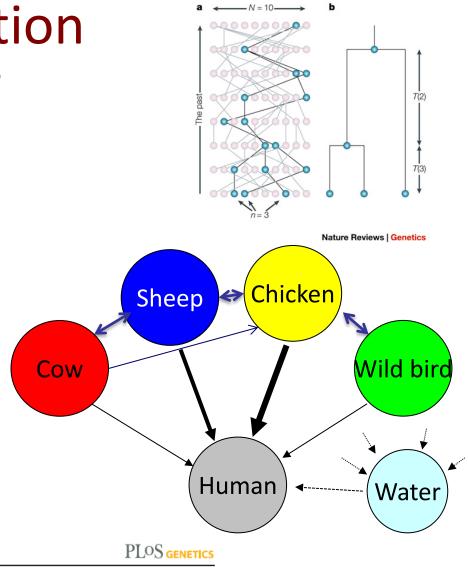
Wagenaar, French and Havelaar, 2013. *Clin Infect Dis, 57, 1600-7* **Preventing Campylobacter at the source: why is it so difficult?**



Mather, Vaughan and French, 2015. *Clin Infect Dis*. 61 S259-65 Molecular Approaches to Understanding Transmission and Source Attribution in Nontyphoidal Salmonella and Their Application in Africa.

Reservoir attribution 'Island model'

- Population genetics / evolution approach
- Reconstruct genealogy
- Used to find out source of human infections
- Flow into the human "island" from animal "islands"



OPEN ORCESS Freely available online

Tracing the Source of Campylobacteriosis

Daniel J. Wilson^{1¤a}*, Edith Gabriel^{2¤b}, Andrew J. H. Leatherbarrow³, John Cheesbrough⁴, Steven Gee⁴, Eric Bolton⁵, Andrew Fox^{4,5}, Paul Fearnhead¹, C. Anthony Hart^{6†}, Peter J. Diggle²

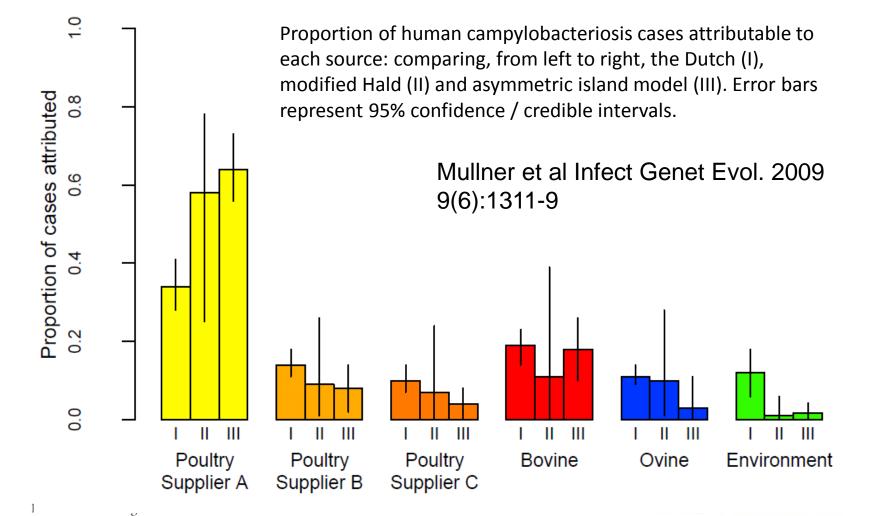


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Reservoir attribution (Mullner et al)



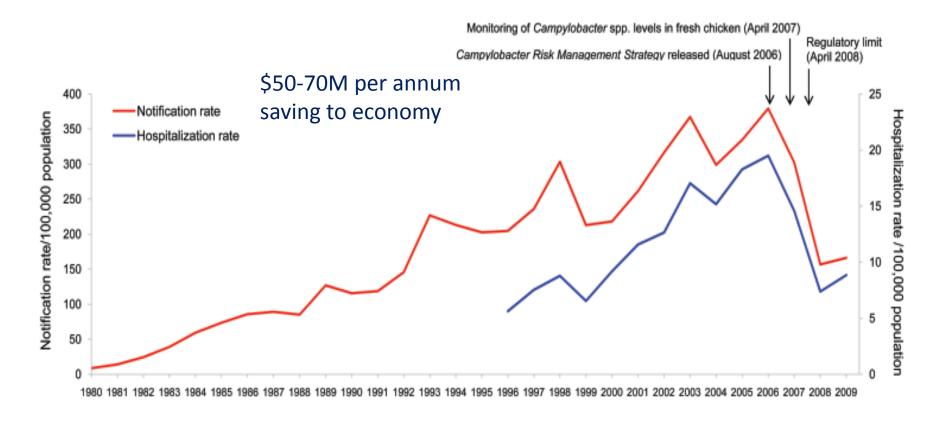




Campylobacter in Poultry – Risk Management Strategy 2007 - 2010



Relationship between campylobacteriosis notifications and hospitalisations (A. Sears)



Data Sources: ESR Ltd notification data; NZHIS hospitalisation data (filtered)

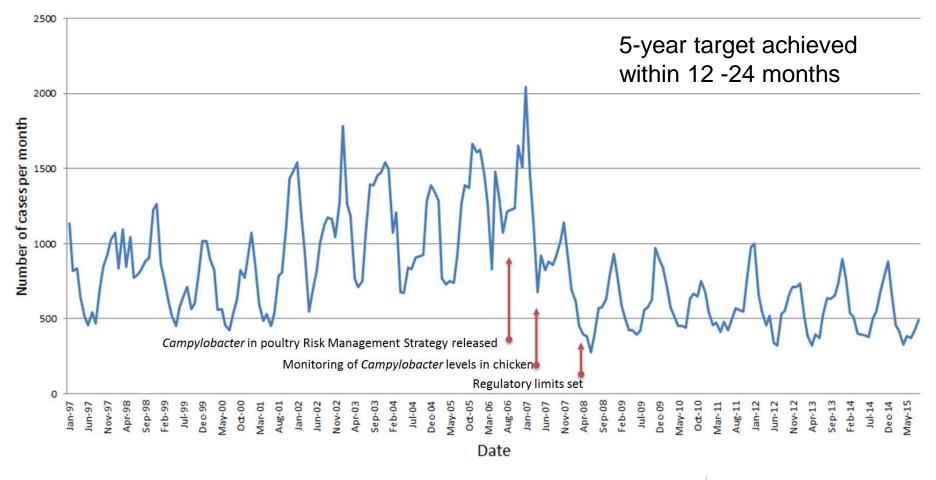


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Marked Campylobacteriosis Decline after Interventions Aimed at Poultry, New Zealand

Sears et al 2011, *Emerging Infectious Diseases* 17, 1007-15

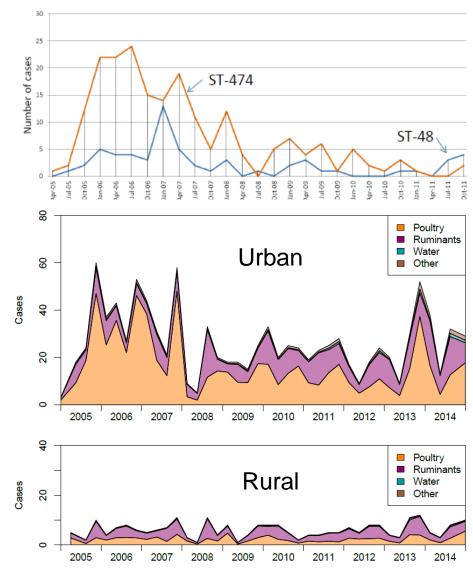
Ann Sears, Michael G. Baker, Nick Wilson, Jonathan Marshall, Petra Muellner, Donald M. Campbell, Robin J. Lake, and Nigel P. French



Surveillance and monitoring: Rise and fall of *C. jejuni* ST 474

- ST-474 found in multiple sources later in epidemic
- Genome sequencing revealed rapid evolution*
- Recombination>mutation
-Waiting for next 'epidemic' strain?

*Biggs et al 2011, PLoS One, 6 (11) e27121 French et al, 2014. Campylobacter Ecology and Evolution, 221-240.



Arrival of ST-6964 and AMR.....



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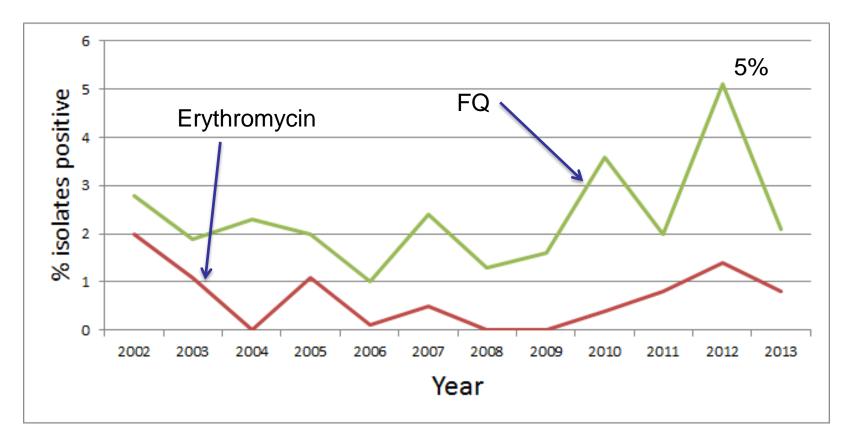
Emergence of ST-6964

(in collaboration with Dr Debbie Williamson, MDU)

- August 2014: first two human cases of new *C. jejuni* ST-6964 detected in Manawatu sentinel surveillance site
- Found in all 3 largest poultry companies
- <u>Resistant to tetracycline and fluoroquinolones</u>
- ST-6964 member of Clonal Complex 354, only other isolates in PubMLST database from China



AMR in Campylobacter in NZ: Human cases to 2013 (data from ESR Ltd)



Low levels of resistance by international standards (UK >25%)

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AMR in Campylobacter in NZ: Poultry

New Zealand Veterinary Journal 58(5), 229-236, 2010

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Scientific Article

Low levels of antibacterial drug resistance expressed by Gram-negative bacteria isolated from poultry carcasses in New Zealand

EJ Pleydell ^{*§} , L Rogers [*] , E Kwan [*] a	and NP French*
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	Disc ^a (µg)	Res (%) ^b	Zone size (mm)														
Drug			≤6	7–15	16	17	18	19	20	21	22	23	24	25	26	27	≥ 28
Erythromycin	15	0.5	1				1			2	3	4	8	19	12	21	122
Ciprofloxacin	5	0											1	1	1	2	188
Enrofloxacin	5	0											1		3	4	185
Nalidixic acid	30	0			2	2	3	8	12	13	19	24	22	33	17	12	26
Chloramphenic	ol 30	0								1	1				9	10	172
Tetracycline	30	0													1		192

^a Concentration of drug within the disc

^b Percentage of isolates with zone sizes within the resistant category for that drug

Very low by international standards



Poultry survey 2015

- Samples:
 - 193 pooled caecal samples
 - 1 week of slaughter
 - 62 farms, 158 sheds



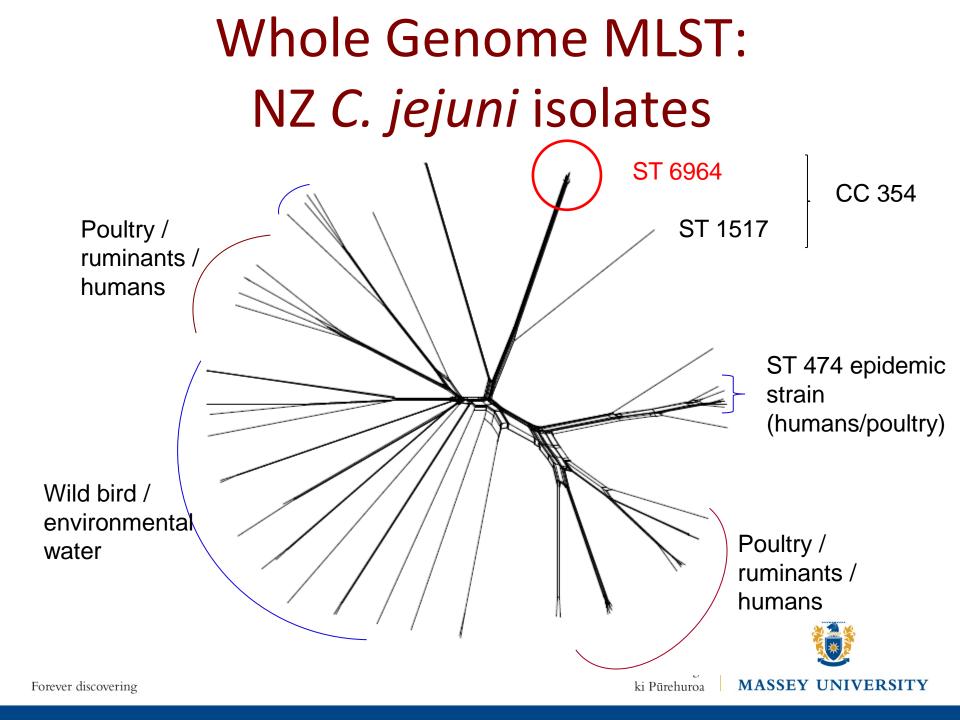
- 37% +ve for ST-6964 tet/cip resistant strain
- 0% +ve on South Island
- All 3 main NI companies +ve (27-62% of samples)
- Also isolated from 'parent' stock



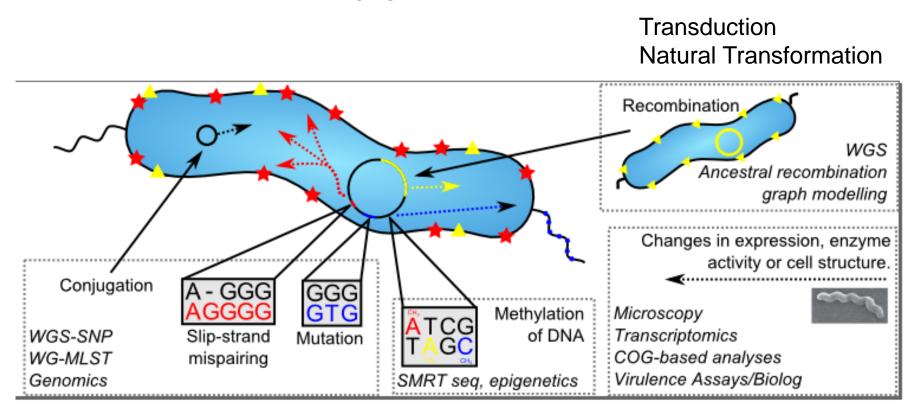
Human survey 2015

- ESR survey (Debbie Williamson and Kirsten Dyet, presented at Antimicrobials 2016, PP2.3)
- May-October 2015, ~300 isolates
- Increased prevalence of FQ and tet resistance
- Most (73%) ST-6964
- Wellington outbreak (chicken liver)



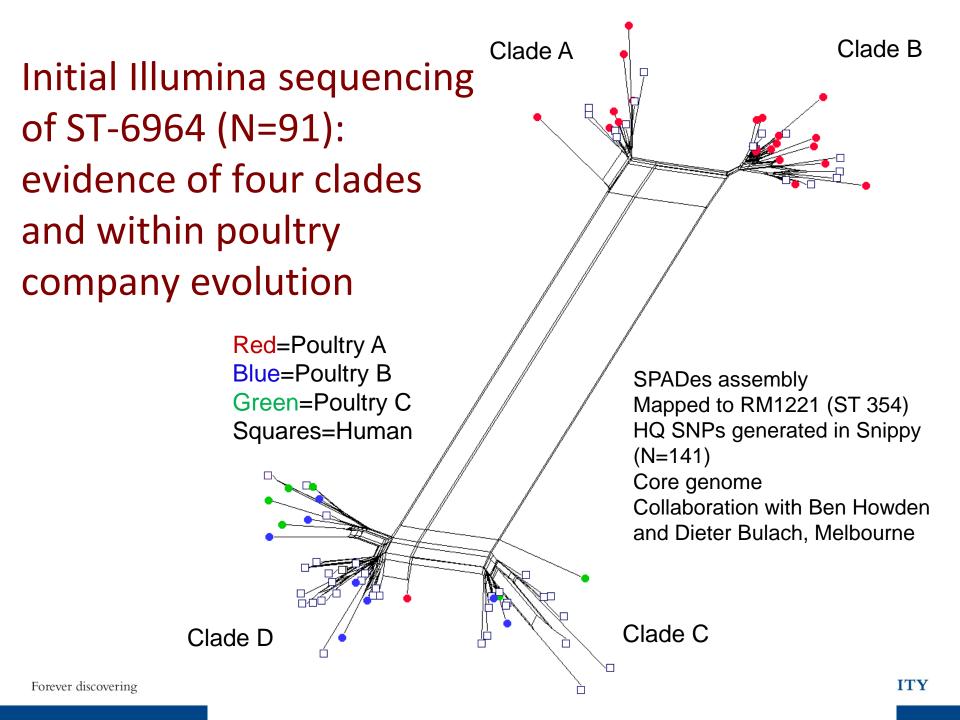


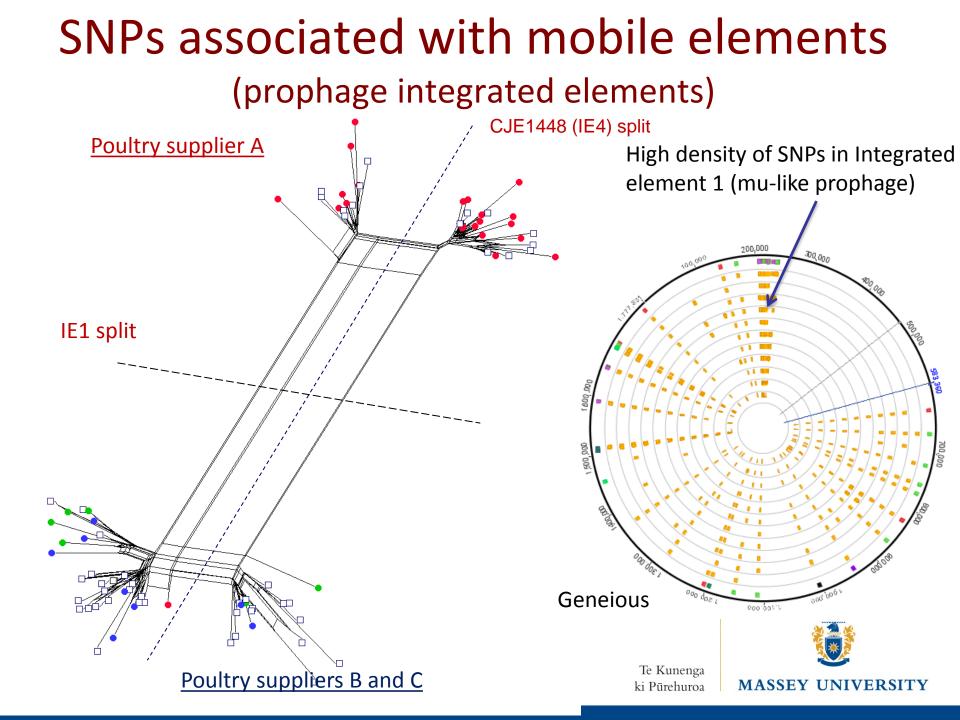
Adaptive mechanisms deployed by Campylobacter



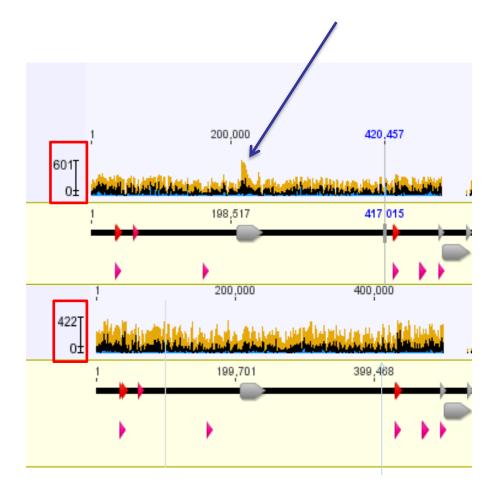
Creates a vast array of possible phenotypes







Ectopic mapping – evidence of extra IE1-like prophage in Supplier A?



IE1

CTT CATA AAC AATC TCTTCTA AA	CTTTG CATTTCTTCAC TG GG CAA AJ	ATA ACT TT A G A G C T	755	218,780	218,800	
			CATANACAATO	TUTTUTAARCTTTG CATTT	CTTCRCTGGGCAAAATAA	CTTT W G A R CTTAATT
100 010 101	242 524	242.4				
466 218,481	218,501	218,	.469	218,494	218,514	218.5
CITCATATACAATCTCTTCTAA,	CTTTG CATTTCTTCAC TG GG CAA AI	ATAACTTTT6AAC1		TOTTOTANG OTTTO CATTT		
Smamamans S	CTTTG CATTTCTTCAC TG GG CAA AJ		C 3 T 3 3 C 3 3 T (TOTTOTANGOTTE CATE		
	CTTTG CATTTCTTCAC TG GG CAAA		CATA		CTTCACTGGGCAAAATAA	
	CTTTG CATTTCTTCAC TG GG CAAA			TOTTOTAA ACTTTG CATTT		
	CTTTG CATTTCTTCAC TG GG CAA A	TTA A CT TT <mark>A</mark> G A <mark>G C</mark> T		TACTTTG CATTT TCTTCTAAACTTTG CATTT	TTCACTGGGCAAAATAA	
CTT CATA AC AATC TCTTCTA A	CTTTG CATTTCTTCAC TG CTTTG CATTTCTTCAC TG GG CAA A		CATAAACAATO	TOTTOTAAGOTTTG CATTT	CTT AATAA	CTTTAG AGATTATTT
	CTTTG CATTTCTTCAC TG GG CAAA					
CTT CATA AC AATC TCTTCTA A				TOTTOTAAACTTTG CATTT TOTTOTAAACTTTG CATTT		CTTTAC ACCTTAATT
CTT CATA AC AATC T	TTG CATTTCTTCAC TG GG CAA A		CATANACAATO	TOTTOTAA CTTTG CATTT	CTTCACTGGGCAAAATAA	CTTT & ACCTTAATT
CTT CATA AC AATC TCTT	TTTCTTCACTG GG CAAA		CATAACAAT	TTG CATTT TCTTCTAAGCTTTG CATTT	CTTCCCCCCAAAATAA	
	CTTTG CATTTCTTCAC TG GG CAA A. CTTTG CATTTCTTCAC TG GG CAA A.			TUTTUTAAGUTTTG CATTT		
	CTTTG CATTTCTTCAC TG GG CAAA			TOTTOTAAGOTTTG CATTT		CTTAATT
	CTTTG CATTTCTTCAC TG GG CAA AI			TOTTOTAA CTTTG CATTT TOTTOTAA CTTTG CATTT		
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	CTTTG CATTTCTTCAC TG GG CAA A			TCTTCTAAGCTTTG CATTT TCTTCTAA CTTTG CATTT		
	ATTTG CATTTCTTCAC TG GG CAA A CTTTG CATTTCTTCAC TG GG CAA A		CATANACAAT	TOTTOTAA ACTTTG CATTT	CTTCACTGGGCAAAATAA	CTTT N G A <mark>G</mark> CTTAATT
	CTTTG CATTTCTTCAC TG GG CAAA			TCTTCTAAGCTTTG CATTT TCTTCTAAGCTTTG CATTT		
	CTTTG CATTTCTTCAC TG GG CAA A			TUTTUTAAGUTTTG CATTT		
	CTTTG CATTTCTTCAC TG GG CAAA			TCTTCTAANCTTTG CATTT		
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	CTTTG CATTTCTTCAC TG GG CAA A CTTTG CATTTCTTCAC TG GG CAA A		CATANACAAT	TUTTUTAA CTTTG CATTT TUTTUTAA CTTTG CATTT	CTTCACTGGGCAAAATAA	CTTT GACCTTAATT
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CTT CATA AC AATC TCT TCTAA	TCTTCAC TG GG CAAA			TCTTCTAAGCTTTG CAT TCTTCTAA <mark>A</mark> CTTTG CATTT		CTTTTG AACTTAATT
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	CTTTG CATTTCTTCAC TG GG CAA A		CATANACAAT	TOTTOTAAGOTTTG CAT	GCAAAATAA	CTTTTG AACTTAATT
	CTTTG CATTTCTTCAC TG GG CAA A. CTTTG CATTTCTTCAC TG GG CAA A		CATANACAATO		CTTCACTGGGCAAAATAA	
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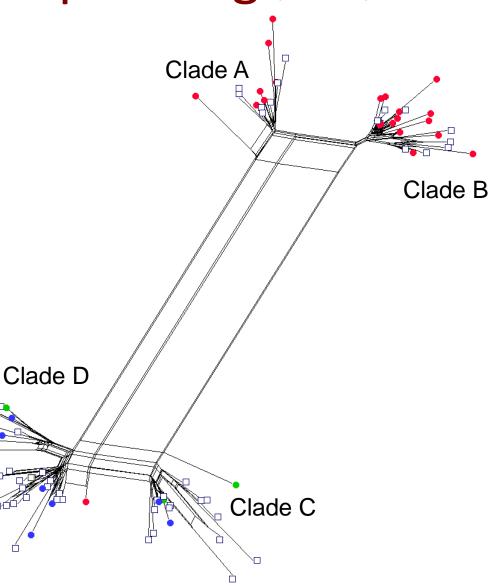
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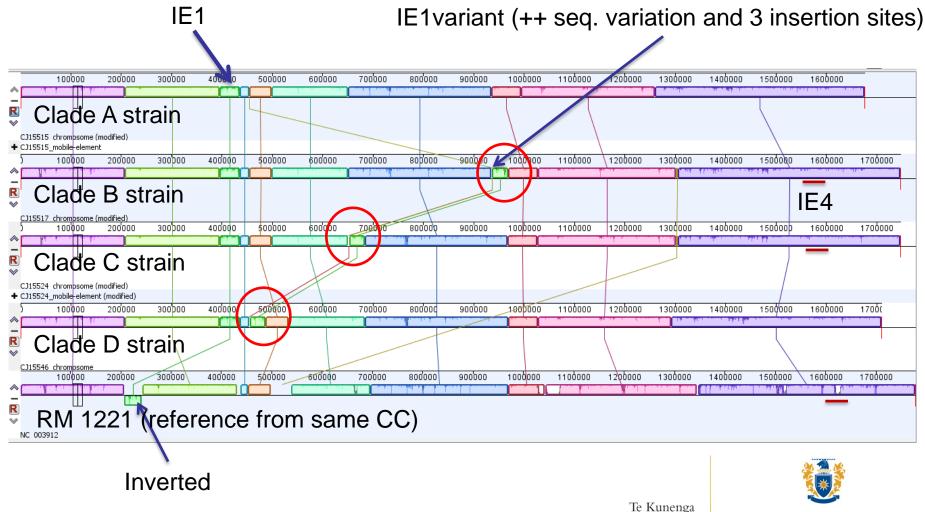
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SMRT (Pacbio) sequencing (MDU)

- Single isolates selected from 4 clades
- Sequenced at MDU, Melbourne (hybrid assembly using Illumina short reads)
- Complete annotated genomes (single contigs)
- Revealed different genome sizes
- Mobile (phage) elements
- Confirmed remnant pTet plasmid in C and D



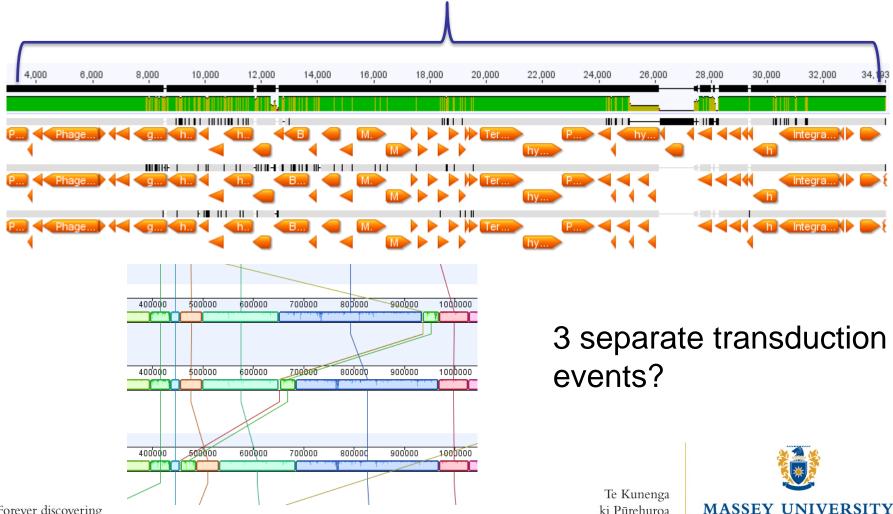
Multiple phage insertions?



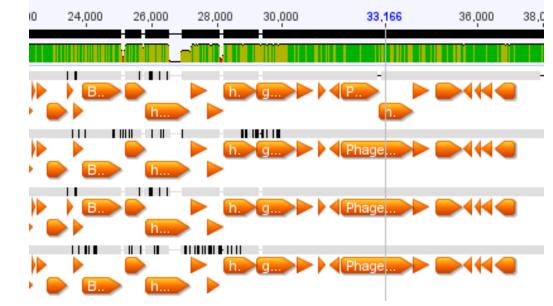
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IE1variant: highly polymorhic phage insertion

IE1v



Number of genes with point mutations in coding regions of core genes low (~10)



Integrated Element 1 (IE1)

Most are in Integrated elements (recombination hotspots)

One recombinant core gene (maf3) motility accessory factor in flagellar (FLAG) region

Consistent with recent common ancestor

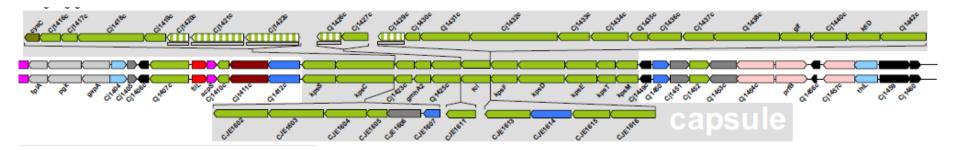


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SSRs and slip-strand mispairing

e.g. Polysaccharide deacetylase – Capsular polysaccharide



High frequency slip strand mispairing On/off switching of genes involved in FLAG, CPS, LOS, OMP, phage resistance 15 SSR g/c and 39 SSR a/t variants identified,

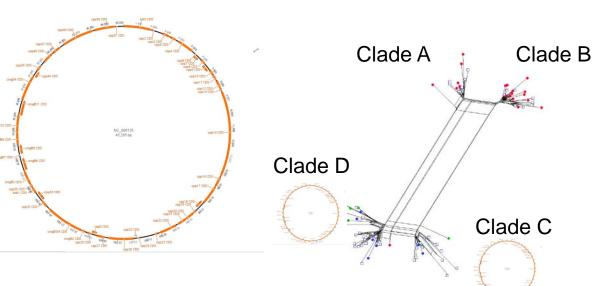


Confirmed *tetO* inserted in all between *kds*B and CJE0905

One of over 100 *tet*O insertion alleles in *C. jejuni* (Cody et al 2015) ST-6964 region close match to strain CG8421 and same insertion site (Poly et al 2012) hypothetical protein CDS Relaxase/mobilization nuclease domain protein CDS putative transposase CDS thrC CDS kdsB CDS Bacterial mobilization protein (MobC) CDS hypothetical protein CDS tetO CDS hypothetical protein CDS vpothetical protein CDS hypothetical protein CDS 834,000 835,000 836.000 837.000 838,000 839.000 840,000 840 761 832,000 833,000 842,00 831,000 tetO gene; tetO dene: gene; aene gene; kdsB gene: kdsB

Remnant conjugative plasmid?

- Absent from poultry supplier A isolates (clades A and B). Present in clades C and D
- Strong homology with other tetO carrying plasmids



hypothetical protein CDS

843,000

fliY CDS

844,000

fliY gene

Key questions?

- Genetic basis for resistance?
 - tetO plasmid and C257T mutation in gyrA
- How long has it been in NZ?
 - Not long??—analysis complicated by SSRs and mobile elements.
- How has it been transmitted between poultry companies?
 - Shared parent and grandparent stock?
 - Feed
 - Complex interrelationships between suppliers (A and C shared parent stock, B and C shared feed)

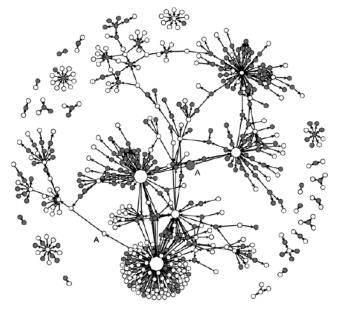


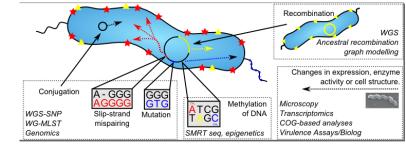
Fig. 2. Social network analysis of feed-related contacts in the New Zealand

Poultry farm feed network (from Lockhart et al 2010)



Key questions?

- What has driven the emergence?
 - FQ use negligible
 - Limited tetracycline use in parent stock
- What is driving evolution?
 - Point mutations? number v. small



- **Natural transformation?** Only one core gene (*maf*3) with evidence of homologous recombination
- Singe Sequence Repeats s ++++ and in previously recognised and other locations
- Conjugation pTet plasmid, tetO chromosomal insertion
- Transduction multiple phage insertions
- Important point: WGS enabled higher resolution attribution to individual poultry supplier



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Conclusions

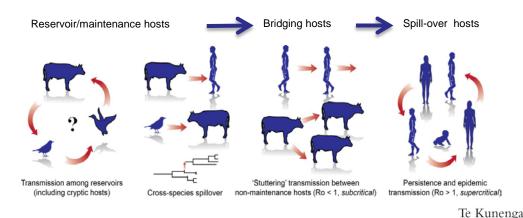
- Improved understanding of epidemiology of enteric zoonoses in New Zealand
 - One Health approach essential (working with Govt and Industry)
 - Application of molecular and genomic epidemiology
 - MLST and Whole genome sequencing models of evolution and transmission
- Source attribution modelling
 - Tools advanced in recent years
 - Applied to *Campylobacter* in NZ and identified food, particularly poultry, most important source, informed policy
 - Major decline in human cases 2007-9
 - New antimicrobial resistant strain emerged in 2014
 - Recent introduction? But high diversity driven by mobile elements.



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Conclusions

- Important to use 'toolbox' approach
 - Space, time, risk factors
 - SA modelling
 - Population genetics/genomics/ecology
- Ongoing and future work
 - Source-assigned case control study (1400 isolates for WGS)
 - Work with poultry industry to reduce prevalence
 - Model development (attribution, ARG models)





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Acknowledgements

- ^mEpiLab team: Patrick Biggs, Jonathan Marshall, Anne Midwinter, Julie Collins-Emerson, Rukhshana Akhter, Charlotte Bolwell, Jackie Benshop, David Hayman, Lynn Rogers, Sam Bloomfield
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- University of Melbourne, MDU Prof Ben Howden, Dr Dieter Bulach
- University of Otago: Prof Michael Baker,
- University of Auckland (Dr Tim Vaughan, Prof Alexei Drummond)
- Prof Paul Fearnhead (Lancaster)
- Prof Martin Maiden, Dr Danny Wilson, Dr Sam Sheppard
- MidCentral Public Health, MedLab Central
- AgResearch A/Prof Adrian Cookson
- MPI Peter van de Logt, Prof Steve Hathaway, Dr Donald Campbell, Dr Craig Thornley
- University of Tasmania: Dr Barbara Holland







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